New Terpenoids from *Amentotaxus* formosana

Hui-Ling Chen,[†] Li-Wen Wang,[†] Huey-Jen Su,[‡] Bai-Luh Wei,[§] Sheng-Zehn Yang,[⊥] and Chun-Nan Lin^{*,†}

School of Pharmacy, Kaohsiung Medical University, Kaohsiung 807, Taiwan, Department of Nursing, Mei-Ho Institute of Technology, Pintung Hsien 912, Taiwan, Institute of Life Science, National Taitung University, Taitung 950, Taiwan, and Department of Forest Resource, Management and Technology, National Pintung University of Science and Technology, Pintung Hsien 912, Taiwan

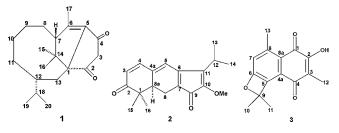
lincna@cc.kmu.edu.tw

Received December 14, 2005

ORGANIC LETTERS 2006

Vol. 8, No. 4 753-756

ABSTRACT



Amentoditaxone (1), possessing an unprecedented diterpenoid skeleton, along with two new terpenoids, amentotaxin WC (2) and amentotaxone (3), were established by extensive analysis of spectroscopic data.

In previous papers, we have reported the isolation and biological activity of constituents from *Amentotaxus formosana*.^{1–5} As part of a continued investigation on the bioactive constituents of Formosan plants, a novel diterpenoid, with a new skeleton, amentoditaxone (1), and two new terpenoids, amentotaxin WC (2) and amentotaxone (3), were isolated from the leaf and heart wood of this plant, respectively. In the present paper, the structure elucidation of these new compounds and the hypothetical biogenetic route for the formation of 1, 2, and 3 are reported.

- [‡] Mei-Ho Institute of Technology.
- [§] National Taitung University.
- $^\perp$ National Pintung University of Science and Technology.
- (1) Su, H.-J.; Day, S.-H.; Yang, S.-Z.; Chiang, M.-Y.; Lin, C.-N. J. Nat. Prod. 2002, 65, 79-81.
- (2) Day, S.-H.; Su, H.-J.; Lin, C.-N.; Yang, S.-Z. Helv. Chim. Acta 2002, 85, 2377–2382.
- (3) Su, H.-J.; Wang, L.-W.; Lin, C.-N.; Day, S.-H.; Wei, B.-L.; Yang, S.-Z.; Won, S.-J. *Helv. Chim. Acta* **2003**, 86, 2645–265.
- (4) Su, H.-J.; Wang, L.-W.; Lin, C.-N.; Day, S.-H.; Wei, B.-L.; Yang, S.-Z.; Won, S.-J. *Helv. Chim. Acta* **2004**, 87, 2723–2725.

10.1021/ol053029m CCC: \$33.50 © 2006 American Chemical Society Published on Web 01/17/2006

The whole plants of *A. formosana* were collected at Kaohsiung Hsien, Taiwan, during July 2001. The CHCl₃ extracts (80 g) of the leaf (3.1 kg) of this plant were chromatographed over silica gel. Elution with *n*-hexane/ acetone (8.5:2.5) yielded **1** (4 mg). The CHCl₃ extracts (6.5 g) of the heart wood (1.6 kg) of this plant also were chromatographed over silica gel. Elution with CH₂Cl₂/ acetone (1:1) and CH₂Cl₂/MeOH (9:1) yielded **2** (5 mg) and **3** (10 mg), respectively.

Compound **1** was isolated as colorless powder, $[\alpha]_D^{25} - 32^\circ$ (*c* 0.075, CHCl₃). The IR spectrum of **1** exhibited an absorption band for the saturated carbonyl moiety (1706 cm⁻¹).⁶ The presence of an unsaturated carbonyl moiety was confirmed by the UV spectrum $[\lambda_{max} 265 \text{ nm } (4.381)].^7$ HREIMS suggested a molecular formula of C₂₀H₃₀O₂ ([M]⁺, 302.2244, Δ -0.0002 mmu).

[†] Kaohsiung Medical University.

⁽⁵⁾ Wang, L.-W.; Su, H.-J.; Day, S.-H.; Tsao, L.-T.; Yang, S.-Z.; Wang, J.-P.; Lin, C.-N. *Planta Med.* **2005**, *71*, 344–348.

⁽⁶⁾ Biemann, K. Tables of Spectral Data for Structure Determination of Organic Compounds; Springer-Verlag: New York, 1989; p 1130.

⁽⁷⁾ Scatt, A. I. Interpretation of the Ultraviolet Spectra of Natural *Products*; Pergamon Press: New York, 1964; p 70.

Table 1. ¹H and ¹³C NMR Spectroscopic Data (δ in ppm, J in Hz) of **1**, **2** (in CDCl₃), and **3** (in CD₃OD)

1			2			3		
position	$\delta_{ m H}$	$\delta_{ m C}$	position	$\delta_{ m H}$	$\delta_{ m C}$	position	$\delta_{ m H}$	$\delta_{ m C}$
		67.8	1		45.6	1		178.8
		207.3	$egin{array}{c} 1 \\ 2 \\ 3 \end{array}$		203.0	$\frac{2}{3}$		170.8
3	α 2.15 d (18.8)	38.0	3	5.96 d (9.6)	124.5	3		109.0
	β 3.23 d (18.8)		4	7.17 d (9.6)	147.8	4		182.7
4		208.8	4a		137.7	4a		137.6
$4\\5\\6$		152.3	5	7.30s	118.7	5		132.9
6		153.3	6		147.9	6		160.1
7	1.87 dd (13.2, 2.8)	47.1	6 7 8		135.0	7	6.67	121.5
8	α 1.77 ddd (21.2, 13.2, 5.2)	23.4	8	α 2.93 dd (14.0, 11.6)	43.2	8		148.7
	$\beta 2.13 \text{ m}$			β 3.11 t (14.0)				
9	α 2.52 ddd (9.6, 5.2, 2.0)	39.0	8a	α 2.93 t (11.6)	43.8	8a		117.2
	$\beta 2.60 \text{ m}$		9		201.1	9		98.0
10	α 1.15 m	33.6	10		147.4	10	$1.76~{ m s}$	25.9
	β 1.33 m		11		142.4	11	$1.76~{ m s}$	25.9
11	α 1.24 m	41.7	12	3.26 m	27.0	12	$1.89~\mathrm{s}$	7.7
	β 1.48 m		13	1.27 d (6.8)	23.5	13	$2.56~{ m s}$	20.6
12	1.15 m	33.7	14	1.27 d (6.8)	23.3			
13	1.56 m	18.3	15	$1.05 \mathrm{~s}$	21.2			
14		44.0	16	$1.31 \mathrm{~s}$	23.5			
15	0.91 s	17.6	OMe	$3.85~\mathrm{s}$	62.0			
16	0.89 s	22.0						
17	1.04 s	33.8						
18	2.60 m	24.9						
19	1.07 d (6.8)	21.4	21.4					
20	1.11 d (6.8)	21.1	21.1					

The structure of **1** was completely assigned by a combination of one- and two-dimensional NMR methods. Its ¹H NMR spectrum (Table 1) contained the signals of two secondary methyls, three tertiary methyls, six methylene groups, and three methine groups. Its ¹³C NMR spectrum (Table 1) further indicated the presence of five methyl, six methylene, three methine, and six quaternary carbons.

Analysis of the ${}^{1}\text{H}{-}{}^{1}\text{H}$ COSY and HMQC experiment for 1 established the connectivities of eight ${}^{1}\text{H}{-}{}^{1}\text{H}$ and ${}^{1}\text{H}{-}{}^{13}\text{C}$ spin systems represented as bold lines (Figure 1). The HMBC correlation of H-12/C-18 and Me-19 and Me-20/C-12 confirmed connectivity between C-12 and C-18. The

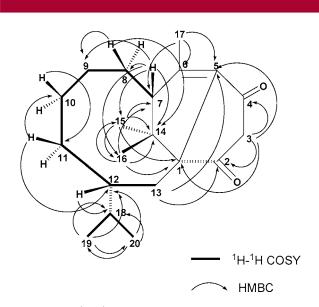


Figure 1. Key ¹H⁻¹H COSY and HMBC correlations of 1.

754

HMBC correlations of H-7/C-8, C-15, C-16 and C-14, Me-15/C-1, C-7, and C-14, Me-16/C-1, C-7, and C-14, Me-15/ C-16, and Me-16/C-15 established the connectivities between C-7/C-14 and C-14/C-1. The HMBC correlations of Me-17/ C-5, C-6, and C-7 and H-13/C-5 and C-2 established the connectivities between C-7/C-6 and C-5/C-6, and C-13 was linked through C-1 to C-5. The HMBC correlations of H-3/ C-1, C-2, and C-4, and C-5, along with the above evidence of IR and UV spectra, confirmed that the β -diketone group was linked between C-4/C-5 and C-1/C-2. Thus, the structure of amentoditaxone was established as **1**.

The relative configurations at C-1, C-7, and C-12 were determined as shown in 1 from the results of a NOESY experiment (Figure 2). A NOESY experiment on 1 showed cross-peaks between H_{β} -8/Me-17, Me-17/H-7, H_{β} -11/H-12, and Me-16/H_{β}-11. The above result suggested that the proton groups at C-7 and C-12 are on the β -side of **1** and the chemical bond between C-1 and C-2 is on the α -side of 1. From the ¹H NMR, COSY, and NOESY spectra, a computergenerated 3D structure of 1 was obtained by using the molecular modeling program, CS CHEM 3D V 3.5.1, with MM2 force-field calculations for energy minimization (Figure 2). The calculated distances between H_{β} -8/Me-17 (2.797 Å), Me-17/H-7 (2.679 Å), H_{β}-11/ Me-16 (1.881 Å), and H_{β}-11/H-12 (3.024 Å) are all less than 4.00 Å. This is consistent with the well-defined NOESY interactions observed for each of these proton pairs.

Compound **2** was isolated as yellowish powder, $[\alpha]_D^{25} + 30^\circ$ (*c* 0.20, CHCl₃). The HREIMS of **2** indicated a [M]⁺ peak at m/z 298.1550, $\Delta -0.0019$ mmu, which corresponded to a molecular formula of C₁₉H₂₂O₃. The IR absorption of **2** implied the presence of a conjugated ketone in a five-membered ring (1718 cm⁻¹), an α,β -unsaturated carbonyl

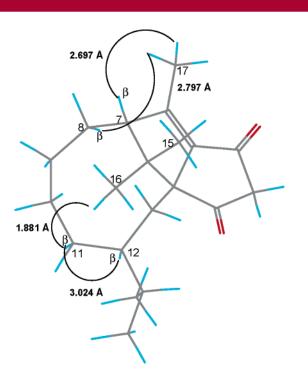


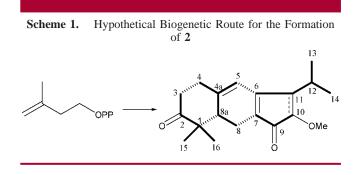
Figure 2. Selective NOESY correlations and relative stereochemistry for 1.

group (1647 cm⁻¹),⁸ and a conjugated C=C moiety (1594 cm⁻¹). The UV spectrum of **2** exhibited absorption maxima at 260 (4.23), 320 (3.55), and 375 (4.06) nm. The ¹H NMR spectrum of 2 (Table 1) showed proton signals for two tertiary methyl groups at δ 1.05 and 1.31, two secondary methyl groups at δ 1.27 (6H, d, J = 6.8 Hz), a methylene group at δ 2.93 (1H, dd, J = 14.0, 11.6 Hz) and 3.11 (1H, t, J = 14.0 Hz), two methine groups at δ 2.93 (1H, t, J =11.6 Hz) and 3.26 (1H, m), a methoxy group at δ 3.85 (s), a *cis*-disubstituted olefin moiety at δ 5.96 (1H, d, J = 9.6Hz) and 7.17 (1H, d, J = 9.6 Hz), and an olefinic proton at δ 7.30 (s). The ¹³C NMR spectrum (Table 1) showed signals for four methyl, a methoxyl, a methylene, two methine, three olefinic carbons, and eight quaternary carbons. The above evidence suggested that 2 contained an α,β -unsaturated carbonyl moiety. The HMBC correlations of H-3/C-4a, H-4/ C-2 and C-8a, H-5/C-6, C-7 and C-8a, Me-15/C-2, C-1, C-8a and C-16, Me-16/C-1, C-2, C-8a and C-15, H_b-8/C-8a, H_a-8/C-4a and H-8a/C-4a, the ¹H-¹H COSY correlation between H₂-8 and H-8a and the NOESY correlations of Me- $16/H_{\alpha}$ -8, H-8a and Me-15 and Me-15/Me-16 suggested that 2 possessed a disubstituted 1,1-dimethyl-8,8a-dihydro-2H,8Hnaphthalene-2-one moiety. The HMBC correlations of H-5/ C-6 and C-7, H₂-8/C-9, MeO-10/C-10, Me-13/C-11, Me-14/C-11, the ¹H-¹H COSY correlations of H-12/Me-13 and Me-14, and the NOESY correlations of H-5/Me-14 and OMe-10/Me-13, with C-9 and C-10 presented as quaternary carbon, confirmed the structure of amentotaxin WC as 2. The NOESY experiment of 2 showed a cross-peak between

 H_{α} -8/Me-16 and Me-16/H-8a and suggested that H-8a and Me-16 are on the α -side of **2**. The presence of a fragment peak at m/z 283 [M - Me]⁺, 269 [M - 2 × Me + H]⁺ and 255 [M - CH(CH₃)₂]⁺ in the EIMS also supported the characterization of **2**.

The HREIMS of **3** gave a molecular peak at m/z 258.0899, Δ -0.0007 mmu, which corresponded to a molecular formula C15H14O4. Its IR spectrum showed absorptions due to hydroxyl (3240 cm⁻¹), carbonyl (1684 and 1635 cm⁻¹), and aromatic (1598 cm⁻¹) functionalities. The UV spectrum of 3 exhibited absorption maxima similar to that of hydroxynaphthaquinones.⁹ The ¹H NMR spectrum of **3** (Table 1) showed proton signals for four tertiary methyl groups at δ 1.76 (6H, s), 1.89 (3H, s) and 2.56 (3H, s), and an aromatic proton at δ 6.67. The ¹³C NMR spectrum (Table 1) showed signals for four methyl, a tertiary aromatic carbon, and 10 quaternary carbons, including two carbonyl carbons. The two tertiary methyl proton signals (Me -3 and -8) in 3 were identical to those of aristolindiquinone, a 3,8-dimethylnaphthaquinone derivative,¹⁰ and the HMBC correlations of Me-12/C-2, C-3 and C-4, Me-10 or Me-11/C-9, Me-10 or Me-11/C-5, H-7/C-5 and C-8a, Me-13/C-7, C-8 and C-8a, and NOESY correlation of H-7/Me-13 suggested that 3 was characterized as 3 or 3a. On the basis of the above result, and the comparison of NMR spectra with those of aristolindiquinone¹⁰ and reported data in the literature,¹¹ it was confirmed that the structure of amentotaxone as 3. The presence of fragment peaks at m/z 243 [M – Me]⁺, 215 [M $- C(CH_3)_2 - H]^+$, 187 [215 - CO]⁺, and 115 [M - b - $Me \times 2 - H$ in the EIMS also supported the characterization of **3**.

Compounds 1, 2, and 3 showed no cytotoxic activity against several human cancer cell lines. Biogenetically, the C-1, C-2, C3, C-15, C-16, C-4, C-4a, C-5, C-8a, C-8, C-6, C-11, C-12, C-13 and C-14 in the skeleton of 2 seem to be derived from isopentenyl diphosphate (Scheme). The C-7, C-9, and C-10 may derive from pyruvate (Scheme 1).¹¹



Therefore, this hypothetical result indicates that Amentotaxus

⁽⁸⁾ Bellamy, L. J. *The Infrared Spectra of Complex Molecules*, 2nd ed.; Methuen & CO LTD: London, 1958; p 149.

⁽⁹⁾ Scott, A. I. Interpretation of the Ultraviolet Spectra of Natural Products; Pergamon Press: London, 1964; p 258.

⁽¹⁰⁾ Che, T.-A.; Ahmed, M.-S.; Kang, S.-S.; Waller, D.-P.; Bingel, A.-S.; Martin, A.; Rajamahendran, P.; Bunyapraphatsara, N.; Lankin, D.-C.; Cordell, G.-A.; Soejarto, D.-D.; Wijesekera, R.-O.-B.; Fong, H.-H.-S. J. Nat. Prod. **1984**, *47*, 331–341.

⁽¹¹⁾ Biemann, K.. *Tables of Spectral Data for Structure Determination of Organic Compounds*; 2nd ed.; Springer-Verlag: New York, 1989; p C120.

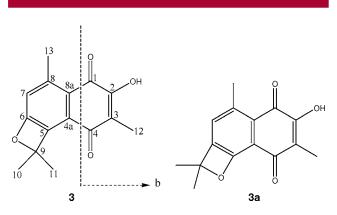


Figure 3. Chemical structure of compounds 3, 3a, and EIMS fragmentation pattern of 3.

formosana uses this hypothetical pathway in the synthesis of new compound **2.**¹² Previously reported terpenoids, amentotaxins WA and WB,¹³ may also use the same biosynthetic pathway. The biosynthesis of **3** agrees with three

head-to-tail isoprenoid units, whereas structure 3a does not. It further supported the characterization of 3. It is valuable to study the biosynthetic pathway of the unprecedented diterpenoid 1.

Acknowledgment. This work was supported by a grant from the National Science Council of the Republic of China (NSC 93-2320-B037-017).

Note Added after ASAP Publication. The version of SI posted January 17, 2006 contained errors. The correct version posted January 26, 2006.

Supporting Information Available: ¹H NMR and ¹³C NMR spectra of **1**. This material is available free of charge via the Internet at http://pubs.acs.org.

OL053029M

⁽¹²⁾ Shigemori, H.; Komaki, H.; Yazawa, K.; Mikami, Y.; Nemoto, A.; Tanaka, Y.; Kobayashi, J. *Tetrahedron Lett.* **1999**, *40*, 4353–4354.

⁽¹³⁾ Day, S.-H.; Su, H.-J.; Lin, C.-N.; Yang, S.-Z. Helv. Chim. Acta 2002, 85, 2377–2382.